Microbially Induced Calcite Precipitation for Sustainable Agriculture and Construction

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Abstract

Microbially induced calcite precipitation is attaining a great importance as 'biomimetic' inspiration. Biomineralization is the process by which living organisms carry out reactions that promote mineral precipitation. Bio inspired engineering is futuristic approach for green civil infrastructure with microbial natural phenomenon. This study was done to evaluate the microbially induced calcite precipitation activity of bacteria on strengthening of soil and building material. Ureolytic strains were isolated and identified using biochemical properties as Alcaligenes sp., Bacillus aeolius, Bacillus naganoensis, Bacillus carboniphilus and Bacillus velezensis by 16srRNA sequencing method. Strains found to have the potential of calcite production were grown on calcite precipitation agar and B4 medium. Characterization of calcite was done using stereomicroscopy, scanning electron microscopy, Fourier Transform Infra-Red Spectroscopy and X-ray diffraction techniques. Both circular and hombohedral shapes of calcites were observed with size ranging between 20 to 600 nm. For the utilization of microbially induced calcite precipitation, activity in soil strengthening, sulfur rich soil (80 ppm) was used for field experiment with Zea mays. Maize grown in soil containing Bacillus velezensis, showed elevated vigor index. Concrete brick ameliorated with Bacillus naganoensis and Bacillus velezensis, are herein reported for first time contributing towards increase in crushing load, compressive strength as well as water absorption capacity in comparison to control and bricks meliorated with other strains. This study has concluded that MICP process can be used for sustainable environment.

Keywords: Microbially induced calcite precipitation, bioconsolidation, biomineralization, biogrouting, ureolytic bacteria, vigor index, *Bacillus velezensis*

1.0 Introduction

1.1 Biomineralization:

It is a process in which bacteria produce minerals like carbonates calcium phosphates etc. The synthesis of minerals categorized into two classes, biologically induced mineralization (BIM) and biologically controlled mineralization (BCM). Minerals are synthesized directly at a specific location within or on the cell only under certain conditions in the case of BCM. In case of BIM, the minerals are formed extracellularly as a result of metabolic activity of the organism (1). Minerals known to be formed by means of BIM through passive surface facilitated mineralization includes phosphates, metal sulfates and carbonates. Among all the minerals that have been associated with biomineralization, carbonates are the most observable. Microbially induced calcium carbonate precipitation (MICCP) can be used in

variety of fields ranging from geotechnology, biotechnology to civil Engineering (1). Biomediated process of soil improvement has been considered as an innovative and new approach in geotechnical engineering that can be used to avoid liquefaction and landslide in loose sand which usually results in foundation deformation, mortality and economical losses (2).

The introduction of Microbiologically Induced Calcite Precipitation (MICP) took the concrete material technology to a different dimension. The deposited inorganic CaCO₃ crystals get attached to the surrounding surfaces. The minute sized bacteria can attach themselves within fine pores and voids; and thereby enhance the structural incorporation of the material. When water seeps through concrete the concrete cracks, it comes in contact with bacteria along with oxygen and calcium-based nutrient, CaCO₃ crystals formation seals the cracks. The precipitation of carbonates by urea hydrolysis using ureolytic bacteria is the most easily controlled mechanism of MICCP (Microbiologically Induced Calcium carbonate Precipitation) along with potential to produce high amounts of carbonates within a short period of time. For the duration of microbial urease activity, 1.0 mol of urea is hydrolyzed intracellularly to 1.0 mol of ammonia (NH₃) and 1.0 mol of carbonate, which instinctively hydrolyzes to form additional 1.0 mol of ammonia and carbonic acid as follows (1).

 \dot{CO} (NH₂) ₂ + H₂ Θ ^{bacteria} NH₂COOH + NH₃)

 $NH_2COOH + H_2O \rightarrow NH_3 + H_2CO_3$

These products equilibrate in water to form bicarbonate, 1.0 mol of ammonium and hydroxide ions that give rise to increase in pH.

 $\begin{array}{c} H_2CO_3 \rightarrow 2H^+ + 2CO_3^{2^-} \\ NH^{3+}H_2O^- \rightarrow NH^{4+} + OH^- \\ Ca^{2+} + CO_2^{-3^-} \rightarrow CaCO_3 \end{array}$

MICP produces calcite and other calcium carbonate minerals like Vaterite or Aragonite (3).

1.2 Biomineralization process in construction:

Concrete is widely used artificial construction material in world. Though it has many advantages such as low price and ability of being cast in any desirable shape, it has some limitations too. Carbonate precipitation has been investigated extensively due to its wide range of technological implication and is a significant aspect of biomineralization. Applications of carbonate mineralization comprises of bioremediation through leaching, solid phase capture of inorganic contaminants or plugging cementation in rock and other porous media fissures and the production of biomimetic materials (4). Urea hydrolysis is the most efficient and the best way of the carbonate producing reactions, as it is easy to control with the potential to produce high concentrations of carbonate for CaCO₃ precipitation (5). Now-a-days innovative materials changed the features of concrete.

2.0 Material and method

2.1 Isolation and identification of ureolytic bacteria

Ureases (urea amid hydrolases, EC 3.5.1.5) are a group of enzymes available in nature amid bacteria, fungi, algae, plants and invertebrates. Hydrolysis of urea (H_2N -CO- NH_2) yields NH_3 and carbonic acid, which is catalyzed by urease enzyme. The Christensen's Urea agar media was used for isolation of ureolytic bacteria from the rhizospheric soil by spread plate method. Urease producing microorganisms convert urea into NH_3 which is alkaline in nature and due to which pink colored colony occur. Urease producing strains were identified based

on the morphological and biochemical tests. Cultures showing more ureolytic activity were identified using 16srRNA sequencing method.

2.2 Determination of urease activity

Urease activity of the isolates was determined using phenol hypochlorite method (6) and electric conductivity method (5).

2.3 Phenol hypochlorite assay method

The urease activity of bacteria was determined by using phenol hypochlorite assay method. Ammonium chloride (50 to 100 μ M) was used as standard. The fermented Christensen's urea broth (250 μ l) was added to the mixture containing 1.0 ml of 0.1 M potassium phosphate buffer (pH 8.0) and 2.5 ml of urea (0.1 M). The mixture was incubated for 5 min at 37 °C. Phenol nitroprusside and alkaline hypochlorite were added, 1.0 ml each and incubated at 37 °C for 25 min. Optical density (OD) was measured at 626 nm and one unit of urease is defined as the amount of enzyme hydrolyzing 1.0 μ mol rea/min (6).

2.4 Electric conductivity method

Urease enzyme activity was carried out by using conductivity method. For enzyme assay, 1.0 ml of fermented Christensen's urea broth was added to 9.0 ml of 1.11 M urea solution. Final conductivity was recorded after 30 min of incubation at 20 0 C by electric conductivity meter (NOEQ 660A, India). Urease activity was represented as mS/cm (millisiemens/centimeter) (5).

2.5 Calcium carbonate precipitation

Calcite precipitation agar (CPA) media was used for screening of bacteria which precipitates CaCO₃ through ureolysis. It contained nutrient broth, 3.0; urea, 20.0; CaCl₂.2H₂O, 28.5; NaHCO₃, 2.12; NH₄Cl, 10.0; agar, 15.0 (g/lit). All the media components were autoclaved, except urea which was membrane sterilized. After autoclaving, urea was added. Bacterial cell suspension was spot inoculated on the CPA plates and kept for incubation at 30 0 C for 48 hr. Stereomicroscopic images were taken for 3-7 days (5).

2.6 Calcium carbonate precipitation in broth

For measurement of $CaCO_3$ precipitation in broth, calcite precipitation broth was supplemented with 2% urea and calcium chloride. The broth was inoculated with 2% inoculum and incubated at 30 0 C for 7 days. Precipitated CaCO₃ was centrifuged; the pellet was dried in oven at 60 0 C for 3 hr. The dried samples were checked for Fourier Transform Infra-Red (FTIR) Spectroscopy (Perkin Elmer, USA)(5).

2.7 Crystal nucleation site development

Tap water (500 ml) + 100 ml (3 M) concentration of filtered urea + 100 ml inoculum were added and incubated for two weeks at 30 $^{\circ}$ C. The deposits of CaCO₃ were centrifuged at 4,300 rpm for 15 min. After centrifugation the deposits were kept inside the hot air oven at 35 $^{\circ}$ C for 6 hr. The dried samples were checked for FTIR, X-ray Diffraction (XRD) Spectroscopy and scanning electron microscopy (SEM) analysis (SEM-Jeol, Tokyo, Japan) (7).

2.8 Concrete brick preparation using calcite producing bacteria

Brick preparation was done according to standard criteria of cement brick. The essential ingredient of concrete block contains cement, aggregate (sand/gravel) and water (Figure 1). These concrete blocks were produced manually. For the production of concrete block, ordinary Portland cement and aggregate (sand/gravel) was mixed in proportion of 1:6. Each brick was made with 2 kg of concrete, 400 g cement (1:6) and enriched with strains which were having the potential to grow in the presence of cement. For control, broth was replaced with water. Bricks were kept undisturbed for 10 days and checked for crushing load and compressive strength using Digital Compressive Testing Machine (2000 KN) (Lawrence and Mayo, India) and water absorption capacity (8). Statistical analysis was done by ANOVA with Tukey HSD using statiskingdom. The experimental results of compressive strength were

compared with control values using ANOVA with Tukey HSD using statiskingdom. A p value of <0.05 was considered as significant.

2.9 Effect of MICP isolates on vigor index of *Zea mays* grown in sulphur rich soil **2.9 (a) Seed germination efficacy of** *Zea mays*

For disinfection, seeds were first kept in 1% mercuric chloride (HgCl₂) for 3 min and then washed with sterile distilled water (DW) for 15 seconds. Five seeds maize were placed on sterile water agar plates and incubated at room temperature (RT).

2.9 (b) Field assay

Sulfur contaminated soil was used to study the effect of biomineralization process on maize crops. Sulfur content of soil was determined using sodium acetate extraction-barium chloride method (Motsara and Roy, 2008). Five flasks of Luria Bertani (LB) [LB broth casein enzymatic hydrolysate, 10.0; yeast extract, 5.0; NaCl, 10.0 (g/lit)] were inoculated with 5 strains and incubated for 7 days at 30 $^{\circ}$ C. Seven squares of 10 x 10 inches were made on the field. Five seeds each of maize were sown into each square and 3-5 g of urea was sprinkled. Enriched LB broths with 5 strains were poured in first five squares. The sixth square was kept as control in which only water was added. The seventh square was kept as control with CaCl₂ and urea. After 30 days, the crops were harvested and vigor index was determined (9). Chlorophyll content was also calculated as per the protocol of Maclachlam (10).

2.10 Determination of soil urease activity

Soil urease activity was done by using the protocol of Tabatabai. The method was based on hydrolysis of urea to NH_3 due to urease activity (11).

3.0. Result and Discussion

3.1 Isolation and identification of ureolytic bacteria

Rhizospheric soil was used for the isolation of ureolytic strains. Out of the 10 isolates, total 5 isolates were found to be ureolytic and further studied for MICP activities. These isolates were identified based on the morphological and biochemical characters (Table 1).

| Tests | Isolates | | | | | | |
|--------------------|-------------------------|---------------------|-------------------------|---------------------------|------------------------|--|--|
| | SSR1 | SSR2 | SSR3 | SSR4 | SSR5 | | |
| Gram character | Gram -ve cocci | Gram +ve rod | Gram +ve rod | Gram +ve short rod | Gram +ve rod | | |
| Motility | Motile | Motile | Motile | Motile | Motile | | |
| Sugar fermentation | | | | | | | |
| i) Glucose | A, G | A, G | - | - | A, G | | |
| ii) Mannitol | A, G | A, G | - | - | - | | |
| iii) Xylose | A, G | A, G | - | - | A, G | | |
| iv) Lactose | - | - | - | - | - | | |
| Nitrate reduction | - | - | - | - | - | | |
| Starch hydrolysis | + | + | + | + | + | | |
| Gelatin | + | + | + | + | + | | |
| Growth at varied | | | | | | | |
| conc. of NaCl (%) | | | | | | | |
| 3 | + | + | + | + | + | | |
| 5 | + | + | - | + | + | | |
| 7 | + | - | - | + | + | | |
| 10 | - | - | - | - | + | | |
| Isolate identified | <i>Alcaligene</i> s sp. | Bacillus aeolius | Bacillus naganoensis | Bacillus carboniphilus | Bacillus velezensis | | |

Table 1: Biochemical characteristics of the isolates:

A: Acid, G: Gas, +: Positive, -: Negative

3.2 16srRNA sequencing method of the microbial isolates

The microbial isolates were identified to be Alcaligenes sp., Bacillus aeolius, Bacillus naganoensis, Bacillus carboniphilus and Bacillus velezensis.

16s rRNA sequence of *Bacillus velezensis* is found to be:

>NMTTAATGCGTTAGCTGCAGCACTAAGGGGGCGGGAAACCCCCTAACACT TAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTCGCTCCC CACGCTTTCGCTCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGG TGTTCCACATCTCTACGCATTTCACCGCTACACGTGGAATTCCACTCTCCTCTTCT GCACTCAAGTTCCCCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGGCTTTCA CATCAGACTTAAGAAACCGCTGCGAGCCTTTACGCCCAATAATTCCGGACAACG CTTGCCACCTACGTATTACCGCGGCTGCTGGCACGTAGTTAGCCGGCTTTCTGGT ACCGTCAAGGTGCCGCCCTATTTGAACGGCACTTGTTCTTCCCTAACAACAGAGC TTTACGATCCGAAAACCTTCATCACTCACGCGGCGTTGCTCAGACTTTCGTCCATT GCGGAAGATTCCCTACTGCTGCCTCCCGTAGGAGTCTGGGCCGTGTCTCAGTCCC AGTGTGGCCGATCACCCTCTCCACCCTCTCAGGTCGGCTACGCATCGTTGCCTTG GTGGTGAGCCGTTACCTCACCTACCAACTAGCTAATGCGCCGCGGGTCCATCTG TGGTAGCCGAAGCCACCTTTTATGTCTGAACCATGCGGTTCAAACAACCATCCGG TTACCCACGTGTTACTCACCCCGTCCGCCGCCGCTAACATCAGGGAGCAAGCTCC CATCTGTCCGCTCGTCCGCATGTATTAGGCACGCCGCCAGCGTTCGTCCTGAGCC AGGA

Based on the report of basic local alignment search tool, bacteria showed 99 % similarity with *Bacillus velezensis*.

3.3 Determination of urease activity

Phenol hypochlorite assay and electric conductivity based method was or urease activity.

3.3.1. Phenol hypochlorite assay

In the medium, the ammonium ions react with the phenol hypochlorite to form greenish blue color indophenol, which absorbs light at 626 nm proportional to initial urea concentration. According to the absorbance of phenol hypochlorite assay, ammonia released during the reaction was calculated by using standard graph of ammonium chloride.

Urea + $2H_2O \xrightarrow{Urease} 2NH_3 + CO_2$ (6)

One mole of urea gets hydrolyzed into two molecules of NH_3 . Hence, according to the graph of enzyme unit verses time of incubation, *Bacillus naganoensis* showed continuous increase in enzyme unit; whereas *Bacillus aeolius* and *Bacillus carboniphilus* showed decreased in enzyme activity after some period of time. *Bacillus velezensis* and *Alcaligenes sp.* showed constant urease enzyme activity (Figure 1).

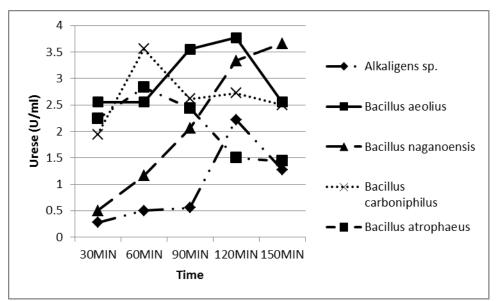
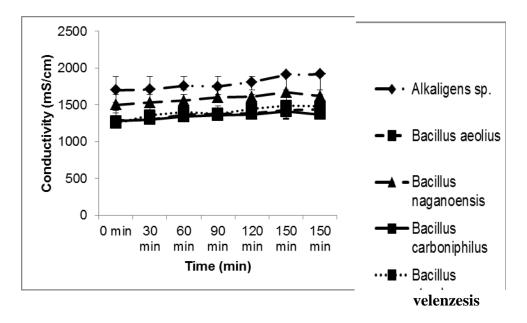


Figure 1: Urease activity of isolates using phenol hypochlorite assay.

3.3.2. Electric conductivity method

Urease assay using conductometer was also done. Once the enzyme was added to substrate, the conductivity was recorded as zero time. Thereafter, it was recorded after 30 min. The urease activity by conductivity is shown in Table 2. The conductivity continuously increased with time in a positive proportional relation with urease activity. It showed stability and consistency in the rate of conductivity. The rate of enzyme activity in *Alcaligenes* sp., *Bacillus naganoensis* and *Bacillus velezensis* found to increase with time. Increased conductivity was due to generation of ionic products of NH⁴⁺ and CO₃²⁻ from non-ionic substrate, urea. The regular increase in conductivity with time is attributed to the regular hydrolysis of urea, which is confirmed by the calculation of correlation coefficients (R) for all tests. The high correlation coefficients indicated strong positive linkage between conductivity and urea hydrolysis (Table 2, Figure 2).

| Isolates | Urease activity by conductivity (mS/cm) against time (min) | | | | | | | Correlation Coefficient | |
|---------------------------|--|------|------|------|------|------|------|----------------------------|--|
| | 0 | 30 | 60 | 90 | 120 | 150 | 180 | (R) | |
| Alcaligenes sp. | 1700 | 1710 | 1760 | 1750 | 1810 | 1910 | 1920 | 95.18 | |
| Bacillus aeolius | 1290 | 1300 | 1370 | 1380 | 1390 | 1430 | 1440 | 0.9654 | |
| Bacillus naganoensis | 1500 | 1530 | 1560 | 1600 | 1610 | 1670 | 1620 | 0.9181 | |
| Bacillus carboniphilus | 1280 | 1300 | 1340 | 1360 | 1370 | 1410 | 1370 | 0.8988 | |
| Bacillus velezensis | 1250 | 1360 | 1400 | 1380 | 1450 | 1490 | 1480 | 0.9279 | |





3.4 Calcium carbonate precipitation on agar plate

Calcite precipitation agar plate was used for CaCO3 precipitation. These isolated colonies showed white precipitation within round the growth area of colonies. On CPA media plates *Alcaligenes sp.* and *Bacillus aeolius* showed rhombohedral crystals of CaCO₃. *Bacillus naganoensis, Bacillus carboniphilus* and *Bacillus velezensis* showed circular crystals with size variations (Table 3, Figure 3).

| Isolates | Agar Medium | | | | | |
|------------------------|----------------------------------|----------------------------------|------------|--|--|--|
| | CPA containing CaCl ₂ | CPA containing CaCO ₃ | B4 | | | |
| Alcaligenes sp. | Rhombohedral | Rhombohedral | Sugar Cube | | | |
| Bacillus aeolius | Rhombohedral | Circular | Oval | | | |
| Bacillus naganoensis | Circular | Small Circular | Circular | | | |
| Bacillus carboniphilus | Small Circular | Large Circular | Circular | | | |
| Bacillus velezensis | Circular | Circular | Circular | | | |

Table 3: Shape of crystals under stereomicroscope grown on different agar medium.

CPA: Calcite precipitate agar, CaCO₃: Calcium carbonate

| Medium Isolates | CPA containing calcium chloride | CPA containing calcium acetate | B4 |
|--------------------|---------------------------------|-----------------------------------|----|
| Alcaligenes sp. | | | |

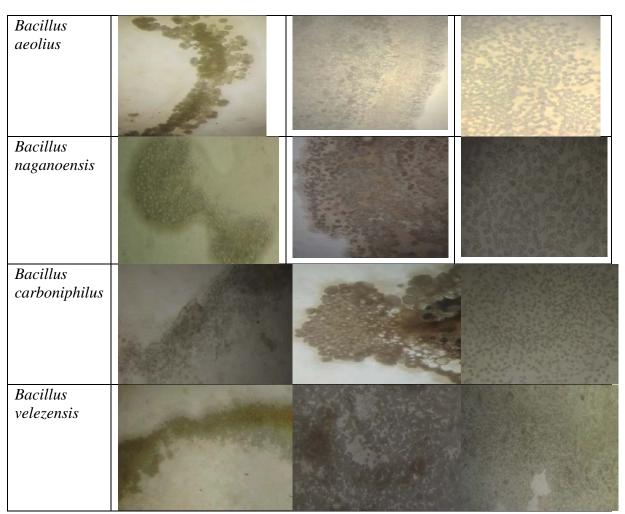


Figure 3: Calcite on calcite precipitation agar and B4 medium under stereomicroscope

3.5 Characterization of crystals

3.5.1. FTIR analysis

The FTIR Spectroscopy was used for qualitative analysis from the characteristic frequencies, which provides information to identify chemical constituents in the compound (Figure 4).

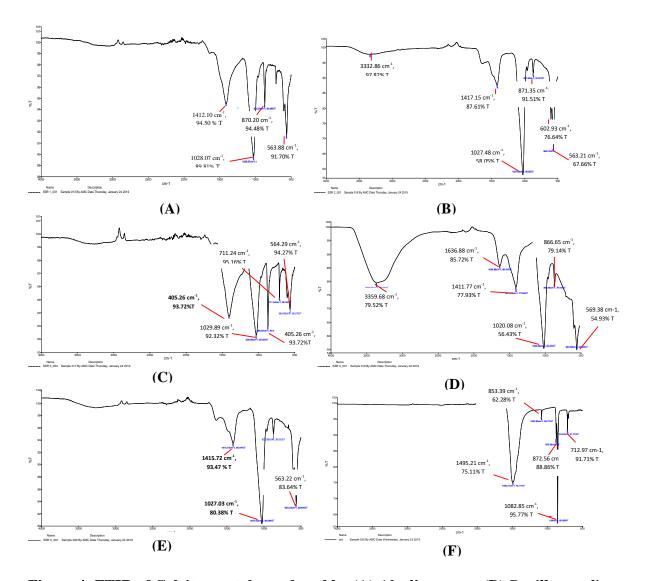


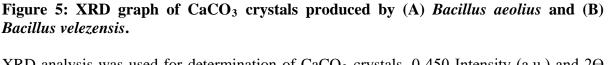
Figure 4: FTIR of Calcite crystals produced by (A) Alcaligenes sp. (B) *Bacillus aeolius* (C) *Bacillus naganoensis* (D) *Bacillus carboniphilus* (E) *Bacillus velezensis* (F) Standard CaCO₃.

FTIR spectra were obtained in regime 4000-550 cm⁻¹. The FTIR spectrum of pure CaCO₃ showed the presence of strong band center around 1495 cm⁻¹. Characteristic of C-O stretching mode of carbonate together with a narrow band around 873 cm⁻¹ of the bending mode. Sharp band at 873 cm⁻¹ is assigned to CaCO₃. Strong band (C-O) at 1444 cm⁻¹ is related with CaCO₃ bond. The (C=O) bond 1415 cm⁻¹ is associated with calcium chloride (12). **3.5.2. XRD analysis**

To ensure that the crystals produced by *Bacillus aeolius* and *Bacillus velenzensis* using crystal nucleation site development method were indeed $CaCO_3$ mineral polymorph, they were examined using x-ray diffraction (Figure 5).

2 Theta (Degree)

ntensity (a.u.)



2 Theta (Degree)

Intensity (a.u.)

XRD analysis was used for determination of $CaCO_3$ crystals. 0-450 Intensity (a.u.) and 2 Θ (Degree) range from 20 to 90⁰ were selected to analyze the crystal orientation. XRD analysis confirms the presence of these multiple mineral forms, including Calcite, Vaterite and Aragonite which is only transiently found in nature. According to XRD pattern, peak was observed at 2 Θ values of 39⁰, 72.35⁰, 29.34⁰, 29.52⁰ in *Bacillus aeolius* whose crystal size ranging from 28-500 nm and in case of *Bacillus velezensis* peak observed at 2 Θ values of 29.52⁰, 43.06⁰, 57.50⁰, 20.78⁰ whose crystal size ranges from 100-630 nm (13).

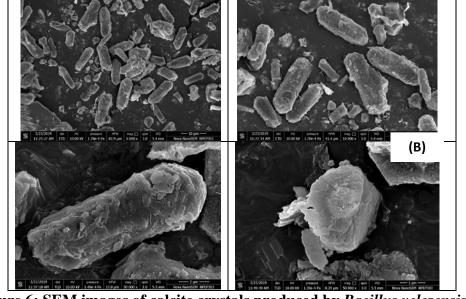
3.5.3. SEM analysis

Carbonate crystals structure were different in size and showed rhombohedral structure of CaCO₃. The SEM image of biocement producing *Bacillus velezensis* on the surface of their cell are shown in Figure 6.

Figure 6: SEM images of calcite crystals produced by *Bacillus velezensis*

3.6 Cement brick preparation using calcite producing bacteria

Out of the five strains, only 3 strains viz, *Alcaligenes* sp., *B. naganoensis* and *B. velezensis* were found to grow and produce calcite in the presence of cementitious mixture. Hence, these 3 strains were used for preparing cement brick which were further tested for preparing



cement brick and also checked for crushing load, compressive strength and water absorption capacity. After checking the crushing load, compressive strength and water absorption capacity of bricks, it was found that the bricks made by using the two organisms *Bacillus naganoensis* and *Bacillus velezensis* are stronger than control. *Bacillus velezensis* as inoculum in cement reparation was found to be more effective as it reduced water absorption capacity and increased compressive strength (Table 4).

| Sample | Weight of Block | Area of Block | Crushing load | Compressive Strength | Water absorption |
|----------------------|--------------------|------------------|------------------|-------------------------|---------------------|
| | (gm) | (mm^2) | (KN) | (N/mm^2) | capacity |
| | | | | | (%) |
| Control (X1) | 2400 | 19089 | 152.2 | 7.97 | 14.00 |
| | | | | | |
| Brick using | 2400 | 16559 | 134.0 | 8.09 | 2.87 |
| Alcaligenes sp.(X2) | | | | | |
| Brick using Bacillus | 2400 | 17679 | 160.0 | 9.05 | 2.10 |
| naganoensis(X3) | | | | | |
| Brick using Bacillus | 2400 | 17084 | 185.0 | 10.83 | 1.96 |
| velezensis (X4) | | | | | |
| | | | | | |

For statistical analysis, the experimental results of compressive strength were compared with control values using ANOVA with Tukey HSD using statiskingdom. A p value of <0.05 was considered as significant. Analysis of variance with Tukey's procedure suggested there were significant differences for the mean based on 2.38198e-12 p-value and 170.290627 F statistic value. The means of the following pairs are proved to be significantly different between x1-x3 (p-value= 2.76834e-7, M= 1.09), x1-x4 (p-value=6.38534e-12, M=2.25), x2-x3 (p-value=0.00000130361,M=0.97), x2-x4(p-value=1.33634e-11, M=2.14), x3-x4 (p-value= 1.01013e-7,M=1.17) pairs using Tukey HSD (honestly significant difference). Increase in the compressive strength suggesting biocement as biodesign material has been reported (14). **3.7 Effect of MICP isolates vigor index of Zea** *mays* **grown in sulphur rich soil**

3.7.1. Seed germination efficacy of Zea mays

After 6 days, the seeds were grown and seed germination efficacy was calculated and found to be 100%.

3.5.2. Field assay

After 30 days, the crops were harvested from the field. Root and shoot length were measured and vigor index was calculated (Table 5). Crops grown in *Bacillus velezensis* treated soil showed significant vigor index (Table 5).

| Treatment | Avg shoot length (cm) <u>+</u> SD | Avg root length (cm) <u>+</u> SD | Chl (mg/g) | | Seed germination (%) | Vigor Index | Urease production (U/ml) |
|---|---|---|---------------|--------|----------------------------|----------------|--------------------------------|
| | | | Chl A | Chl B | | | |
| Alcaligenes sp. + urea+ CaCl ₂ | 9.3±0.4 | 13.7±0.4 | 0.0236 | 0.0026 | 100 | 930 | 2300 |

| Table 5: | Chlorophyll | content and | vigor | index of | plants: |
|----------|-------------|-------------|-------|----------|---------|
| | | | | | |

| Bacillus | 9.4±0.4 | 16.1±0.4 | 0.0018 | 0.0018 | 100 | 940 | 2550 |
|-------------------------|---------|--------------|--------|---------|-----|-----|------|
| aeolius + | | | | | | | |
| urea+ $CaCl_2$ | | | | | | | |
| Bacillus | 8.1±0.4 | 15±0.4 | 0.0008 | 0.00008 | 100 | 810 | 2310 |
| naganoensis | | | | | | | |
| + urea+ | | | | | | | |
| CaCl ₂ | | | | | | | |
| Bacillus | 8.6±0.4 | 14.4 ± 0.4 | 0.003 | 0.0003 | 100 | 860 | 2300 |
| carboniphilus | | | | | | | |
| + urea+ | | | | | | | |
| CaCl ₂ | | | | | | | |
| Bacillus | 7.6±0.4 | 19.1±0.4 | 0.002 | 0.002 | 100 | 760 | 2670 |
| velezensis + | | | | | | | |
| urea+ CaCl ₂ | | | | | | | |
| Control | 7.1±0.4 | 14.3±0.4 | 0.0012 | 0.0012 | 100 | 710 | 2620 |
| (Water) | | | | | | | |
| Control | 7±0.4 | 11.67±0.4 | 0.0014 | 0.0014 | 100 | 700 | 1867 |
| (CaCl ₂ + | | | | | | | |
| urea) | | | | | | | |

SD: Standard deviation

3.8 Urease activity of soil

Bacillus naganoesis and Alcaligenes sp. showed less urease activity, whereas *Bacillus aeolius, Bacillus carboniphilus* and *Bacillus velezensis* treated soil showed higher urease activity. Solidification of sand is dependent upon urease distribution and its activity which is important to avoid erosion (15). Relationship between calcite production and urease activity has been reported in mutated strain of *Sporosarcina pasteurii* for its applicability as construction structure to fill the gaps (6).

4.0 Conclusion

- Concrete brick ameliorated with *Bacillus naganoensis* and *Bacillus velezensis* can have application in construction i.e., in the making of buildings. These concrete bricks will be cheap and have good properties in comparison to the cement bricks. These microorganisms are reported for the first time in making of the concrete bricks.
- The microorganisms *Alcaligenes* sp., *Bacillus aeolius, Bacillus naganoensis, Bacillus carboniphilus* and *Bacillus velezensis* showing calcite precipitation were found to increase the plant growth. So, these microorganisms can be used for sustainable agriculture which will be eco-friendly and economical.

Competing Interest

The authors declare that no competing interest exists among them.

Ethical statement

Authors declare that there is no violation of ethical regulations occurred during their work.

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